Borrelia burgdorferi Requires the Alternative Sigma Factor RpoS for Dissemination within the Vector during Tick-to-Mammal Transmission

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Abstract

While the roles of rpoSBb and RpoS-dependent genes have been studied extensively within the mammal, the contribution of the RpoS regulon to the tick-phase of the Borrelia burgdorferi enzootic cycle has not been examined. Herein, we demonstrate that RpoS-dependent gene expression is prerequisite for the transmission of spirochetes by feeding nymphs. RpoS-deficient organisms are confined to the midgut lumen where they transform into an unusual morphotype (round bodies) during the later stages of the blood meal. We show that round body formation is rapidly reversible, and in vitro appears to be attributable, in part, to reduced levels of Coenzyme A disulfide reductase, which among other functions, provides NAD⁺ for glycolysis. Our data suggest that spirochetes default to an RpoS-independent program for round body formation upon sensing that the energetics for transmission are unfavorable.


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Introduction

Bacterial genomes typically encode multiple alternative sigma factors that reversibly associate with the RNA polymerase apoenzyme to promote the transcription of specific subsets of genes in response to changing environmental conditions [1]. One of the best studied alternative sigma factors is RpoS, the master regulator of the general stress response in E. coli (RpoSEc) [2,3]. Regulation of rpoSEc is multifaceted, occurring at the transcriptional and translational levels, as well by proteolysis [2,3]. Induction of the RpoSEc regulon occurs in response to a variety of stressors, including low nutrient availability, high osmolarity, reactive oxygen intermediates and low pH [4]. Up to 10% of the genome may be regulated either directly or indirectly by RpoSEc with the composition of the regulon determined by the specific stressor [5]. RpoS orthologs exist in diverse Proteobacteria, including numerous pathogenic species, and have been shown to control genes involved in a wide range of adaptive processes [6]. For example, RpoS is essential for biofilm formation in Pseudomonas aeruginosa [7], growth and survival of Legionella pneumophila within phagolysosomes [8], and infectivity of mice by Salmonella enterica [9].

B. burgdorferi (Bb), the Lyme disease spirochete, uses just three sigma factors, σ⁴⁰, RpoN, and RpoS, to modulate its transcriptome during its enzootic cycle. Transcription of rpoSBb is directly activated by the alternative sigma factor RpoN (BB0450) with the aid of two enhancer proteins: the response regulator Rap2 (BB0763) and the DNA-binding protein and Fur orthologue, BosR (BB0647) [10–12]. Additional levels of control are provided by the small RNAs hfq (bb0268) and dsrA (bb0577) as well as the carbon storage regulator csrA (bb0184) [12]. Based upon micro-array analysis of mammalian host-adapted spirochetes [13], it was proposed that RpoS functions as a gatekeeper for controlling the upregulation of mammalian host-phase genes (e.g., ospC) and the downregulation of tick-phase genes (e.g., ospA). By quantitative real time PCR (qRT-PCR), we established that rpoS is induced during the nymphal blood meal but is not expressed by spirochetes within replete larvae or unfed nymphs, thereby delineating the RpoS ON (fed nymph) and OFF (fed larvae and unfed nymph) states during the tick phase of the enzootic cycle [13,14]. Expression patterns of RpoS-dependent genes in the tick, however, reflect more than a simple ON/OFF bipartite regulation. For example, OspA is induced within the midgut during nymphal feeding but not by all spirochetes [14–16]. In addition, OspA continues to be expressed at high levels in the midgut after engorgement and is “OFF” only after organisms have completely mammalian host-adapted [14,16–18]. Within the mammal, RpoS-dependent genes are required to establish infection and presumably continue to be expressed until spirochetes are acquired by naïve larvae [12,14,19,20].

While the roles of rpoSBb and individual RpoS-dependent genes have been studied extensively within the mammal [21], the contribution of the RpoS regulon to the tick-phase of the enzootic cycle has yet to be examined. In this study, we defined the
contours of RpoS-dependent gene expression within larval and nymphal life stages and demonstrate, for the first time, that RpoS is required for the transmission of Lyme disease spirochetes by feeding nymphs. Although RpoS-deficient spirochetes replicate at wild-type levels during the nymphal blood meal, mutant organisms display an aberrant morphology, previously referred to as round bodies or cysts [22–24], as feeding progresses. We show that round body formation is rapidly reversible, and in vitro bodies or cysts [22–24], as feeding progresses. We show that round body formation is rapidly reversible, and in vitro appears to be attributable, in part, to reduced levels of Coenzyme A disulfide reductase, which among other functions, provides NAD⁺ for glycolysis. Our findings reveal a previously unsuspected role for RpoS in the physiological adaptations required for tick-to-mammal transmission as well as the existence of a proposed default survival mode (round body formation) triggered when spirochetes are unable to produce sufficient energy for dissemination within the nymph.

Results

Lyme disease spirochetes require RpoS-dependent genes other than ospC to traverse the nymphal midgut

We began by devising an experimental strategy to test our hypothesis that one or more genes within the RpoS regulon are required for the dissemination of spirochetes during nymphal feeding. While it has long been known that ospC is induced during nymphal feeding [15], there has been controversy as to whether this lipoprotein functions in the tick or the mammal [19,25,26]. The strategy we devised also enabled us to clarify the site of OspC function. Because spirochetes lacking either RpoS or OspC are avirulent by needle inoculation [26,27], we used the immersion method [28] to introduce wild-type (WT, CE162), ΔrpoS (CE174), and ΔospC (CE303) isolates into naïve larvae (Table 1); larvae infected by immersion feeding were allowed to molt into nymphs that then were fed on naïve mice. As described in Materials and Methods and summarized in Table 2, we cultured (i) hemolymph at 72 h post-placement to monitor penetration through the midgut; (ii) skin excised from the bite site at 24, 48 and 72 h post-repletion to assess penetration through the salivary glands and transmission; and (iii) distant tissues at 2 and 4 weeks post-repletion to assess dissemination within the mouse. WT spirochetes were isolated from hemolymph at 72 h post-placement, from the bite site at 48 and 72 h post-repletion and from multiple tissues at 2 and 4 weeks post-repletion. The ΔrpoS isolate was not recovered from either hemolymph or distal sites, while the ΔrpoS-complemented isolate (CE467) was recovered from both. In contrast to ΔrpoS organisms, ΔospC Bb were recovered from hemolymph at 72 h post-placement and the bite site but only at 48 h post-repletion. Importantly, the time frames during which WT and ΔospC organisms were recovered from hemolymph cultures were highly similar in each experiment (n = 3; data not shown), suggesting that the cultured samples contained comparable numbers of organisms. Given that RpoS is essential for the stress responses of many bacteria [2,6], it is possible that spirochetes lacking this sigma factor were unable to survive within the midgut during feeding. qPCR and semi-solid plating, however, revealed virtually identical burdens of WT, ΔrpoS, and ΔospC isolates in unfed and replete nymphs; furthermore, similar incubation times were required for the recovery of WT and ΔrpoS organisms by semi-solid plating (Figure S1; p ≥ 0.05). Comparable results were obtained using fluorescent ΔrpoS mutant and complemented strains described in subsequent sections. Thus, B. burgdorferi requires one or more RpoS-dependent genes to traverse the midgut, while OspC functions exclusively within the mammal.

Table 1. Borrelia burgdorferi strains used in this study.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Description</th>
<th>Antibiotic Resistance¹</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>CE162</td>
<td>Wild-type 297 clone</td>
<td>NA</td>
<td>[27]</td>
</tr>
<tr>
<td>CE174</td>
<td>CE162 ΔrpoS; non-fluorescent rpoS mutant</td>
<td>Erm</td>
<td>[27]</td>
</tr>
<tr>
<td>CE303</td>
<td>CE162 ΔospC; non-fluorescent ospC mutant</td>
<td>Kan</td>
<td>This study</td>
</tr>
<tr>
<td>CE467</td>
<td>CE174 ΔrpoS/CE320; non-fluorescent RpoS-complement</td>
<td>Erm and Kan</td>
<td>[36]</td>
</tr>
<tr>
<td>Bb914</td>
<td>CE162 with PflaB-gfp in cp26; fluorescent wild-type</td>
<td>Gent</td>
<td>[30]</td>
</tr>
<tr>
<td>Bb1058</td>
<td>CE174 with Pmut-gfp in cp26; fluorescent rpoS mutant</td>
<td>Gent and Erm</td>
<td>This study</td>
</tr>
<tr>
<td>SE186</td>
<td>Bb1058 ΔrpoS/pcE320; fluorescent RpoS-complement</td>
<td>Gent, Erm, and Kan</td>
<td>This study</td>
</tr>
<tr>
<td>CE56</td>
<td>CE162 ΔrpoS/pcE320; fluorescent flaB transcriptional reporter</td>
<td>Kan</td>
<td>[59]</td>
</tr>
<tr>
<td>CE103</td>
<td>CE162 ΔospA/pcE320; fluorescent flaA transcriptional reporter</td>
<td>Kan</td>
<td>[36]</td>
</tr>
<tr>
<td>CE309</td>
<td>CE162 Δcdr; non-fluorescent</td>
<td>Erm</td>
<td>[29]</td>
</tr>
<tr>
<td>CE1655</td>
<td>CE309 Δcdr/pcE323; non-fluorescent CadR-complement</td>
<td>Erm and Kan</td>
<td>[29]</td>
</tr>
</tbody>
</table>

¹ Antibiotic resistance determined by growing spirochetes in the presence of the following antibiotics: erythromycin (Erm, 0.06 μg/ml); kanamycin (Kan, 400 μg/ml); and/or gentamycin (Gent, 50 μg/ml).
doi:10.1371/journal.ppat.1002532.t001
RpoS-dependent gene expression by spirochetes in fed nymphs differs from that of mammalian host-adapted organisms

The above results established the importance of defining the RpoS regulon during the tick-phase of the enzootic cycle. At the present time, however, delineation of the spirochete’s transcriptome within ticks is not technically possible. Therefore, we used qRT-PCR to measure transcript levels for a panel of 10 RpoS-upregulated and 6 RpoS-repressed tick-phase genes derived from our microarray analysis of WT and ΔrpoS spirochetes cultivated within díalysate membrane chambers (DMCs) [13]. Representative results are shown in Figure 1, while results for the remainder of the panel are presented in Figure S2. All eight genes previously shown to be absolutely RpoS-dependent in DMC-cultivated spirochetes were induced during nymphal feeding with little to no transcript detectable in either fed larvae or unfed nymphs. Two of the upregulated *Borrelia* genes in our panel (*bb0728/cdr* and *bb0670/chw3*) were selected because they were transcribed by both RpoS and ΔrpoS [13]. In accord with these findings, transcripts for both genes were significantly enhanced in fed compared to unfed nymphs. It was interesting to note, however, that in fed larvae, when RpoS is OFF, both *cdr* and *chw3* were expressed at levels comparable to those in fed nymphs. Particularly noteworthy, all six tick-phase genes repressed by RpoS in the mammal were well expressed during the nymphal blood meal; in fact, three (ospA, *bb0365*, and *bb0329*) were expressed at their highest levels within fed nymphs.

To corroborate the above expression data, we also examined the same panel of *Bb* genes in fed nymphs that had been infected as larvae by immersion with either WT-*gfp* (*Bb914*) or ΔrpoS-*gfp* (*Bb1058*) isolates. Little to no transcript was detected for 8 of the 10 RpoS-upregulated genes in fed nymphs infected with ΔrpoS-*gfp* organisms compared to their WT-*gfp* counterparts (Figures 1A and S2A). Expression of *cdr* and *chw3* was reduced significantly in nymphs infected with the mutant compared to WT, establishing definitively that these genes are partially-dependent on RpoS for expression during the nymphal blood meal [Figure 1B and [29]]. Consistent with results obtained using naturally-infected ticks, the genes repressed by RpoS in mammalian host-adapted organisms were expressed at equal or higher levels by ΔrpoS-*gfp* spirochetes compared to WT-*gfp* (Figures 1C and S2B).

Confoocal microscopy of WT spirochetes harboring fluorescent transcriptional reporters for *flagB* or *ospC* was performed as an additional means of confirming that RpoS-repressed tick-phase genes are expressed throughout the nymphal blood meal. As shown in Figures 1D and 1E, the overall density and distribution of the two reporter strains within fed nymphal midguts were indistinguishable. Collectively, our findings indicate that genes whose expression is upregulated by RpoS are likely to be expressed both during and after transmission by nymphs while RpoS-dependent repression appears to occur only after spirochetes have fully adapted to the mammalian host.

### Spirochetes introduced into larvae by immersion undergo biphasic dissemination during nymphal feeding

Lyme disease spirochetes disseminate through the midguts of naturally-infected nymphs in two stages: an initial adherence-mediated migration phase during which non-motile spirochetes advance as networks toward the basolateral surface of the epithelium, followed by a transition into motile organisms at or near the basement membrane which then access the hemocoel [30]. The data in Table 2 suggested that spirochetes lacking RpoS are defective in one or both of these phases. In order to examine the contribution of RpoS to biphasic dissemination, we first established that transmission of WT-*gfp* *Bb* by nymphs infected as larvae by immersion recapitulates the sequence of events previously described for naturally-infected nymphs [30]; a detailed schematic indicating how confocal images of unfed and fed midguts were acquired is presented in Figure S3. Spirochete burdens in unfed and fed nymphs infected by immersion were comparable to those detected in nymphs infected by the natural route (Figure S4A: p = 1.000). By confocal microscopy, the distribution of spirochetes within the midguts of unfed and feeding nymphs infected by either method were highly similar (Figure S4B and [30]). The kinetics of transmission by nymphs infected by either route, based on the recovery of organisms in hemolymph, also were indistinguishable (Table 2 and [30]).

### Table 2. One or more RpoS-dependent gene products, independent of *ospC*, are required for spirochete’s to penetrate into the hemolymph; see Figure S1 for spirochete burden analyses.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Description</th>
<th>Hemolymph</th>
<th>Bite Site</th>
<th>Distal Tissue Sites</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>a,b,c</td>
<td>24 h</td>
<td>48 h</td>
</tr>
<tr>
<td>CE162</td>
<td>WT</td>
<td>21/25</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Bb914</td>
<td>WT-*gfp</td>
<td>35/37</td>
<td>0/24</td>
<td>8/20</td>
</tr>
<tr>
<td>CE303</td>
<td>ΔospC</td>
<td>31/34</td>
<td>0/20</td>
<td>7/20</td>
</tr>
<tr>
<td>CE174</td>
<td>ΔrpoS</td>
<td>0/39</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Bb1058</td>
<td>ΔrpoS-*gfp</td>
<td>0/56</td>
<td>0/24</td>
<td>0/24</td>
</tr>
<tr>
<td>CE467</td>
<td>ΔrpoS+*rpoS</td>
<td>22/25</td>
<td>ND</td>
<td>ND</td>
</tr>
</tbody>
</table>

*Hemolymph was collected from feeding nymphs 72 h post-placement and cultured in BSK-II. The denominator represents the total number of ticks analyzed.

†Cultures were monitored for the presence of spirochetes by dark field microscopy for 8 weeks.

The time frames in which organisms (WT, WT-*gfp*, ΔospC, ΔrpoS+*rpoS*, and ΔrpoS-*gfp+*rpoS*) were recovered from the hemolymph was highly similar in each experiment (n = 3).

‡Skin of a C3H/HeJ mouse (4–6 sites per mouse) was excised from the site where a nymph was attached at the indicated time post-repletion. A minimum of 3 mice were tested per isolate. The denominator represents the total number of bite sites analyzed; ND = not determined.

*Ear punches were performed at 2 and 4 weeks post-feeding and cultured in BSK-II.

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**Requires RpoS for Tick Transmission**

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**Spirochetes introduced into larvae by immersion undergo biphasic dissemination during nymphal feeding**

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Figure 1. Contours of the RpoSBbb regulon in I. scapularis. qRT-PCR analysis of (A) absolutely and (B) partially RpoS-dependent upregulated genes and (C) RpoS-repressed genes selected from microarray data derived from Bb cultivated within DMCs [13]. A representative sample of genes is shown; data for the remaining genes are presented in Figure S2. Expression profiling was performed using fed larvae, unfed and fed nymphs that had been naturally-infected with WT Bb as well as fed nymphs that had been infected as larvae by immersion with either WT-gfp or ΔrpoS-gfp isolates.
DrpoS organisms are confined to the midgut lumen and develop into an aberrant morphotype during nymphal feeding.

Having confirmed the suitability of the immersion method, we proceeded to characterize DrpoS spirochetes within nymphal midguts. For these experiments, we generated fluorescent versions of our previously characterized DrpoS mutant CE174 (DrpoS-gfp; Bb1058) and complemented strain (DrpoS-gfp+p; SE186) (Table 1 and Figure S5). At the outset, we confirmed using qPCR and semi-solid plating that DrpoS-gfp Bb, like its non-fluorescent counterpart, replicated to wild-type levels within feeding nymphs (Figure S1B) but was unable to traverse the midgut or establish infection (Table 2). As expected, the complement was fully virulent by tick-and needle-inoculation (Tables 2 and S2).

By confocal microscopy, the distribution of DrpoS-gfp Bb in midguts isolated from unfed nymphs did not differ from that of WT-gfp organisms (Figure 2A; also see Figure S3). By 48 h post-placement, however, there was a marked reduction in the overall density of DrpoS-gfp Bb on the luminal surfaces and between epithelial cells (Figure 2B). In addition, many of the mutant organisms located between cells were thin, ragged and blebbing (Figure 2B, ‘midway’ panel). At 72 h, the difference between midguts harboring WT-gfp and DrpoS-gfp isolates was striking (Figure 3). Whereas WT-gfp organisms formed typical confluent networks (Figure 3A; WT-gfp), DrpoS-gfp Bb were rarely observed (Figure 3B, DrpoS-gfp ‘coronal’), a result ostensibly discordant with the comparable spirochete burdens detected in fed nymphs infected with either strain (Figures S1B and S1C). We conjectured that DrpoS-gfp organisms were confined to the lumen at the late feeding time point but that the blood meal was interfering with their visualization. We confirmed this assumption by carefully removing a portion (~30–40%) of the blood meal from isolated 72 h-fed midguts and acquiring transverse optical sections from distal portions of diverticula to reduce light scattering and absorption by the luminal contents (Figures S3B–3D). As shown

![Figure 2. Spirochetes lacking RpoS are distributed normally within unfed nymphal midguts but are destroyed between epithelial cells early during feeding.](https://www.plospathogens.org/article/content/figure/1002532/002/5232/8/2_5232_002_002.png)
in Figure 3B (ΔrpoS-gfp, transverse panel), the lumens of ΔrpoS-gfp-infected midguts were, in fact, crowded with organisms. Epifluorescence microscopy of intact and cryosectioned midguts (Figures 4B and S6B, respectively) and silver-staining of paraffin-embedded specimens (Figures 4E and 4H) also revealed lumens packed with ΔrpoS-gfp spirochetes at the 72 h time point. Importantly, complementation restored the ability of ΔrpoS-gfp Bb to form networks and extend to the basement membrane; see also Figure S5 for complementation verification by protein profile analysis. The leftmost images in each panel depict the full composites (basement membrane to luminal surface) of the midgut, while 3-μm composite images show spirochetes at the luminal surface, midway through the epithelial layer, and at the basement membrane. Numbers in lower right hand corner indicate the optical depth of the image; scale bars = 25 μm. Representative cryosections of 72 h-fed midguts infected with WT-gfp and ΔrpoS-gfp isolates are presented in Figure S6.

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Figure 3. RpoS is required to form networks during nymphal feeding. (A) At 72 h post-placement, WT-gfp organisms form networks that encase midgut epithelial cells and extend to the basement membrane. (B) ΔrpoS-gfp Bb are not observed in intact midguts imaged in the ‘coronal’ plane but are visualized within the lumens of midguts that had been punctured to remove a portion of their contents and imaged by acquiring transverse optical sections from distal portions of the diverticula to reduce light scattering and absorption by the blood meal. A detailed schematic illustrating how coronal and transverse confocal images were acquired is presented in Figure S3. (C) Complementation restores the ability of ΔrpoS-gfp Bb to form networks and extend to the basement membrane; see also Figure S5 for complementation verification by protein profile analysis. The leftmost images in each panel depict the full composites (basement membrane to luminal surface) of the midgut, while 3-μm composite images show spirochetes at the luminal surface, midway through the epithelial layer, and at the basement membrane. Numbers in lower right hand corner indicate the optical depth of the image; scale bars = 25 μm. Representative cryosections of 72 h-fed midguts infected with WT-gfp and ΔrpoS-gfp isolates are presented in Figure S6.

Loss of RpoS and CoADR exacerbate the formation of round bodies in vitro

Spirochetes cultivated under nutrient-limiting conditions form morphotypes [22–24,31,32] resembling the round bodies observed in nymphal midguts. These reports led us to hypothesize that RpoS-deficiency creates a metabolic lesion that promotes the formation of round bodies during the nymphal blood meal. To test this idea, we took advantage of the observation by Alban et al. [32] that spirochetes form round bodies during a relatively short incubation period in RPMI. As shown in Figure 7, the percentage of WT-gfp round bodies increased steadily over 4 days, reaching a plateau of ~50% on day 3; a significantly higher percentage of ΔrpoS-gfp spirochetes formed round bodies on days 1 through 4 (Figure 7, p<0.002 for each day). Complementation of ΔrpoS-gfp significantly reduced the percentages of round bodies to levels lower than those observed with WT-gfp on days 3 and 4 (Figures 7 and S7, p<0.01 for both days). Moreover, we confirmed by TEM that round bodies formed by WT-gfp and ΔrpoS-gfp isolates in RPMI were identical to each other (Figure 6B) and closely resembled those in nymphal midguts (Figure 6A). Previously, Xu...
et al. [33] proposed that ΔospC spirochetes have a defective membrane surface. It was thus surprising to find that the percentage of ΔospC round bodies was actually lower than WT levels at days 3 and 4 and that the rate of round body formation was comparable to that observed for WT-gfp organisms (data not shown).

In Bb, the enzyme CoADR, encoded by cdr, contributes to the regeneration of NAD⁺ for glycolysis and the maintenance of the pool of Coenzyme A for biosynthetic functions [34,35]. Our finding that transcription of cdr, one of the few metabolic genes within the RpoS regulon, is markedly reduced in ΔrpoS-gfp Bb during nymphal feeding (Figure 1B) suggested that diminished levels of CoADR might be involved in the enhanced formation of round bodies by ΔrpoS-gfp in RPMI. As shown in Figure 7, the percentages of round bodies formed by Δadr and ΔrpoS-gfp Bb were not significantly different on days 1 and 2 (p>0.6 for both days); by days 3 and 4, however, the rate of round body formation by Δadr organisms significantly outpaced both WT and ΔrpoS spirochetes (p=0.0006 and p<0.002 for WT and ΔrpoS, respectively, on both days). As with ΔrpoS-gfp Bb, complemented Δadr spirochetes formed round bodies at levels lower than those of WT-gfp on days 3 and 4 (Figures 7 and S7, p<0.002 on both days).

Round bodies elongate rapidly upon addition of BSK-II

Alban et al. [32] noted that round bodies formed in RPMI elongate within seconds following the addition of normal rabbit serum (NRS). We sought to confirm this observation, which suggests that neither the metabolic lesion that triggers round body formation nor the resulting rearrangement of the spirochete’s protoplasmic cylinder involve alterations in the cell envelope that require extensive biosynthetic repair. In our hands, the addition of NRS did not promote round body recovery. However, the addition of an equal volume of Barbour-Stoenner-Kelly medium (BSK-II), either with or without NRS, induced the rapid (~3 s) recovery of WT-gfp and ΔrpoS-gfp round bodies (Video S1). Interestingly, one or two highly refractile inclusions often could be observed within round bodies formed in RPMI by both Bb isolates; these structures remained prominent even after bacteria had fully elongated. Based on visualization of 32 round bodies formed by

Figure 4. Spirochetes lacking RpoS are confined to the lumen and develop into round bodies during the later stages of feeding. (A–C) Representative epifluorescence images of midguts isolated at 72 h post-placement containing (A) WT-gfp, (B) ΔrpoS-gfp or (C) RpoS-complemented isolates; midguts appear white when imaged by dark-field microscopy. (D–I) Representative silver-stained paraffin-embedded sections of midguts isolated from 72 h-fed nymphs infected with (D and G) WT-gfp, (E and H) ΔrpoS-gfp or (F and I) RpoS-complemented isolates. Insets in (G) and (H) are enlargements of the boxed regions showing WT-gfp spirochetes and ΔrpoS-gfp round bodies, respectively; scale bars: A–C = 50 μm; D–F = 25 μm; and G–I = 10 μm.

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WT-gfp and ΔrpoS-gfp Bb (16 per isolate) in RPMI, we found that each recovered into a single spirochete (Video S1).

To further the comparison between round bodies formed within ticks and RPMI, we next investigated whether round bodies within midguts isolated at 72 h post-placement also recover upon the addition of BSK-II. Indeed, as shown in Figure 8A, tick-derived ΔrpoS round bodies elongated into spirochetes within minutes. Importantly, the addition of RPMI did not promote recovery (Figure 8B).

Discussion

Studies involving RpoS Bb have focused primarily on the contribution of RpoS-dependent genes to virulence in mammals [21]. The role of RpoS Bb during the tick-phase of the enzootic cycle, on the other hand, has been largely unexplored despite multiple lines of evidence suggesting that genes controlled by this alternative sigma factor might function during tick-to-mammal transmission [12,13,36]. In this study, we demonstrate that ΔrpoS spirochetes remain confined to the midgut throughout the blood meal, and, thus, are unable to progress beyond the initial stage of dissemination. Unexpectedly, RpoS deficient spirochetes replicating within the midguts of feeding nymphs undergo a dramatic morphologic transformation comparable to that induced by nutritional deprivation in vitro. The high degree of phylogenetic divergence between RpoS Bb and its orthologs in Proteobacteria [37], the ostensible lack of overlap between the RpoS regulons in Bb and E. coli [13], and the finding that RpoS Bb does not protect spirochetes against “conventional” external stressors (e.g., reactive oxygen species, low pH, high salt) [27] previously led us to conclude that RpoS Bb is not a central regulator of a general stress response [13]. The findings presented here, however, argue that RpoS-mediated alterations in the spirochete's transcriptome include physiologic adaptations required for dissemination within the vector.

ospC is widely regarded as the prototypical RpoS-dependent gene [12]. While there is universal agreement that OspC is essential for the ability of Bb to establish murine infection, there is lingering controversy as to whether this surface lipoprotein functions within the vector or the mammal [38]. Pal et al. [25] reported that spirochetes lacking OspC were unable to penetrate the salivary glands of feeding nymphs. Rosa and colleagues [19,26], on the other hand, visualized ΔospC organisms within salivary glands and demonstrated that OspC was required for survival within mice following needle-inoculation. Our data, derived entirely by tick-inoculation, are in accord with those of the Rosa group. We found that spirochetes lacking OspC were recovered from hemolymph and reached the tissue surrounding the bite site with kinetics indistinguishable from those of WT Bb but were cleared from the skin within 48–72 h post-repletion, a time frame which closely mirrors that previously reported using needle-inoculation [26]. The inability of ΔrpoS Bb to traverse the midgut, therefore, cannot be attributed to the lack of OspC. We recently reported that spirochetes lacking cdr possess a growth defect within feeding nymphs and are avirulent by needle inoculation [29]. Thus, RpoS-dependent genes fall into three

Figure 5. ΔrpoS Bb form round bodies within nymphal midguts during the later stages of feeding. Representative TEM images of nymphal midguts containing WT-gfp and ΔrpoS-gfp spirochetes isolated at (A–B) 48 or (C–D) 72 h post-placement. Color overlays are used to highlight normal spirochetes (red), round bodies (blue), and peritrophic membranes (PM, green); scale bars = 2 μm.

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broad categories: (i) ospC-like genes that are required solely within the mammal; (ii) genes, such as *bb0365* [39,40], that promote dissemination within the tick; and (iii) genes, such as *cdr*, that function in the arthropod as well as the mammal [29].

The qRT-PCR studies described herein brought to light a striking difference in the transcriptional profiles of WT *Bb* within larval and nymphal ticks as compared to mammalian host-adapted (i.e., DMC-cultivated) organisms: while genes upregulated by RpoS in DMCs also were induced in fed nymphs, genes repressed by RpoS in mammals were expressed at comparable levels by *Bb* in all tick life stages. A comparison of transcript copy numbers in WT and *ΔrpoS* Bb demonstrated that several tick-phase genes (i.e., *ospA*, *bb0240*, *bb0365*, and *bbA52*) are, in fact, subject to some degree of downregulation during the nymphal blood meal. One possible mechanistic explanation for the limited RpoS-mediated repression observed within fed nymphs is that signals produced during the blood meal activate regulatory pathways that oppose the downregulation of tick-phase genes during transmission. The Hk1/Rrp1 two-component system, which promotes tick-adaptation during feeding but is not required for mammalian infection [41–43], is a candidate counter-regulatory pathway. Our findings add to a large body of evidence that the “classical” paradigm for reciprocal regulation of tick- and mammalian host-phase genes (*ospA*/*ospC* being the prototypes) is valid only within the mammal [13,14,16,44,45]. A number of recent studies have helped to elucidate the biological significance for the prolonged transition from the tick- to the mammalian host-adapted state [46–51].

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**Figure 6.** Wild-type and *ΔrpoS* round bodies are structurally similar both within feeding nymphs and under nutrient-limiting conditions *in vitro*. Representative TEM images of *ΔrpoS-gfp* and WT-*gfp* spirochetes and round bodies in (A) nymphal midguts isolated at 48 or 72 h post-placement or (B) *in vitro*-derived organisms following incubation in BSK-II or RPMI for 3 days. In panels A and B, arrows indicate examples of protoplasmic cylinders; scale bars = 500 nm.

doi:10.1371/journal.ppat.1002532.g006
Discernible differences between WT-

and

rpoS

in vitro-derived organisms. (A) The addition of BSK-II induced the recovery of round bodies into elongated spirochetes. (B) Round bodies do not recover when midguts were submerged in RPMI. Scale bars = 10 μm. See also the Video S1 of the rapid recovery of in vitro-derived organisms.

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Figure 7. Loss of RpoS and CoADR exacerbates round body formation in vitro. Bb were incubated in RPMI for 1–4 days. A minimum of 300 organisms were counted per strain for each time point. Experiments were performed in triplicate; error bars represent means ± SEM. Representative images of fields used to quantify round body formation are shown in Figure S7. The percentages of round bodies formed by organisms were counted per strain for each time point. Experiments were performed in triplicate; error bars represent means ± SEM. Representative

images of fields used to quantify round body formation are shown in Figure S7. The percentages of round bodies formed by 

isolates were significantly greater than WT on days 1 through 4 (p<0.002). Round body formation by 

and 

isolates was significantly different on days 3 and 4 (p<0.002).

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Figure 8. Round bodies within fed nymphal midguts recover into elongated spirochetes. 

were removed at 72 h post-placement and midguts dissected into RPMI. (A) The addition of BSK-II induced the recovery of round bodies into elongated spirochetes. (B) Round bodies do not recover when midguts were submerged in RPMI. Scale bars = 10 μm. See also the Video S1 of the rapid recovery of in vitro-derived organisms.

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Figure 8. Round bodies within fed nymphal midguts recover into elongated spirochetes. 

were removed at 72 h post-placement and midguts dissected into RPMI. (A) The addition of BSK-II induced the recovery of round bodies into elongated spirochetes. (B) Round bodies do not recover when midguts were submerged in RPMI. Scale bars = 10 μm. See also the Video S1 of the rapid recovery of in vitro-derived organisms.

The most striking aspect of the 

phenotype, the transformation from typical spirochetes to round bodies, did not become apparent until 48–72 h post-attachment. During this late stage of feeding, the nutritional requirements of midgut epithelial cells immensely increases as the cells switch from undifferentiated reserve cells into hyperactive digestive cells, while spirochetes replicate extensively around them [30,56]. Presumably, an intense competition for blood meal nutrients ensues at this interface. Within this same 24 h window, a small percentage of organisms that reach the basement membrane become motile, a transition that enables them to access the hemocoel but likely also substantially adds to their energy requirements. Spirochetes lacking RpoS replicate to WT levels within the midguts of feeding nymphs and exhibit growth kinetics equivalent to those of WT as demonstrated by qPCR and semi-solid plating. These findings argue strongly that the formation of round bodies by 

cannot be attributed to their inability to utilize the blood meal for biosynthetic purposes but rather suggest that loss of RpoS creates a rapidly reversible metabolic lesion that provokes the structural rearrangements that give rise to round bodies. The ability of 

round bodies derived from tick
midguts to recover within minutes after the addition of BSK-II (i.e., without replication) strongly supports this contention. To shed light on this unusual morphotype, we turned to an in vitro system in which WT Bb transform into round bodies during several days of incubation in RPMI [52]. Although the culture conditions obviously differ considerably from the nutrient-replete midgut, the round bodies formed by WT and ΔrpoS Bb in vitro closely resemble those observed in feeding nymphs. Several other features of this in vitro system are particularly instructive and support its utility for elucidating the in vivo morphotype. First, round bodies elongate rapidly in the presence of BSK-II, indicating that their cell envelopes are intact. Second, spirochete numbers do not increase in this system, demonstrating that round body formation is divorced from replication. Third, and most importantly, compared to WT organisms, ΔrpoS Bb exhibited a pronounced tendency to form round bodies, suggesting that the metabolic lesion in feeding nymphs is operative in vitro. This supposition led us to examine clo, whose enzymatic product, CoA-thr, provides the cofactor NAD⁺ for glycolysis as a byproduct. While the clo phenotype provides proof of principle that an energy-related gene product is involved in round body formation, additional genes within the RpoS regulon also may contribute to this phenomenon.

A fundamental question posed by this work is whether round body formation is a unique stress response to nutrient limitation or an aberration of the laboratory with no role in the natural ecology of the spirochete. Formation of round bodies by WT spirochetes argues for its biological relevance as does the sheer complexity of the process, its elegant recovery mechanism, and the stress-related circumstances in which it is observed. Round body formation by WT spirochetes in RPMI can be conceptualized as a response to inadequate energy generation. In contrast, round body formation by WT spirochetes within the midguts of feeding nymphs is sporadic; perhaps the competition between spirochete and epithelial cells for micro-nutrients in the blood meal is particularly intense in localized regions of the midgut. What is clear is that, in both settings, physiologic adaptations mediated by RpoS inhibit round body formation. This formulation implies that (i) the apparatus for round body formation and recovery is poised to operate within the background during feeding but is suppressed either directly or indirectly by an RpoS-dependent gene product and (ii) spirochetes default to round bodies upon sensing that the energetics for transmission (i.e., transition to motility) are unfavorable. Underlying many studies of round body formation in the literature has been the presumption that its primary importance relates to persistence within the mammal [22,23,24]. We propose that round body formation has evolved to support the tick phase of the cycle and predict that there are circumstances, as yet undefined, when spirochetes within the tick resort to this survival program on a large scale in order to maintain a population of transmissible organisms.

Materials and Methods

Ethics statement

All animal experimentation was conducted following NIH guidelines for housing and care of laboratory animals and in accordance with University of Connecticut Health Center regulations after review and approval by the UCHC Institutional Animal Care and Use Committee.

Culture and maintenance of bacterial strains

*B. burgdorferi* isolates used in these studies (Table 1) were cultivated in modified BSK-II [57] supplemented with 6% rabbit serum (Pel-Freeze Biologicals). Selection and maintenance of *B. burgdorferi* strains was performed in media supplemented with the appropriate antibiotics (Table 1). The plasmid content of all isolates was monitored as previously described [58]. Standard temperature-shift experiments and growth curves were performed as previously described [13]. *Eubacterium coli* strains were maintained in Luria-Bertani broth (LB) [1% tryptone, 0.5% yeast extract, 1% NaCl] with the appropriate antibiotic. Selection was performed on LB agar plates (LB with 1.5% agar) supplemented with the appropriate antibiotic.

DNA manipulations and routine cloning

Routine molecular cloning and plasmid propagation were performed using *E. coli* Top10 cells (Invitrogen). Routine and high-fidelity PCR amplification reactions were performed using Choice Taq (DenVille Scientific) and Takara ExTaq (Fisher Scientific), respectively. Plasmid DNAs were purified using Qiagen Midi and Spin Prep kits. Nucleotide sequencing was performed by Agencourt Bioscience Corp.

Strain construction

Strains CE162, CE174, CE467, CE56, CE103, Bb914, CE309, and CE1655 have been previously described [27,29,30,36,39] (Table 1). To generate an ospC deletion mutant, we amplified overlapping upstream and downstream regions containing ospC (bbs19) and flanking sequences from strain 297 using primers BBB18#1F/BBB19#1R(NgoMIV) and BBB19#1F (NgoMIV)/BBB22#1R, respectively (Table S1). The resulting amplicons were cloned into pCR2.1-TOPO (Invitrogen), re-amplified using the same primers as above, and gel-purified. Purified upstream and downstream amplicons were mixed 1:1 and amplified for 4 cycles with Ex Taq in the absence of primer to promote annealing/extension of the overlapping regions followed by 30 cycles in the presence of BBB18#1F and BBB22#1R flanking primers. The joined amplicon was subsequently cloned into pCR2.1-TOPO, digested with EcoRI, and subcloned into similarly digested pUC19 to yield pCE216. A Plr-kan cassette conferring resistance to kanamycin was amplified from pTA-KanG [60] using PflgB(NgoMIV) and KanR(BamHI) and cloned into pCR2.1-TOPO. The Plr-kan cassette was liberated by digestion with NgoMIV and BamHI and ligated into NgoMIV-BglII digested pCE216 to yield pCE217. Approximately 10 μg of gel-purified ospC::kanR insert, released from pCE217 by digestion with NgoMIV and BamHI, was electro-transformed into CE162 (Table 1) as previously described [61]. A transformant (CE303) was selected in BSK-II medium containing the appropriate antibiotic and subsequently confirmed by PCR to be an ospC deletion mutant using the BBB18#1F (extd) and BBB22#1R primers (Table S1). Bb1058, the fluorescent rpoS mutant, was generated by electro-transforming CE174 with pMC1916 [30] followed by selection in BSK-II containing erythromycin (0.06 μg/ml) and gentamicin (5 μg/ml). Inactivation of rpoS was confirmed by PCR using the primers PrpoS-5#1F and RpoS-3XbaI (Table S1). To generate the fluorescent ΔrpoS complement, competent Bb1058 cells were transformed with a wild-type copy of rpoS obtained on pCE320 as previously described [27]. A transformant, SE186, was selected in BSK-II medium containing kanamycin, erythromycin, and gentamicin and subsequently confirmed to contain rpoS by PCR using the primers rpoS-prom(extd) and rpoS-3′XbaI (Table S1). Complementation was confirmed by the production of OspC and DbpA following temperature-shift as assessed by silver-stain and immunoblot, respectively, as previously described [27].
Assessment of *Bb* infectivity by needle inoculation

Infectivity was assessed using 4–6 week old female C3H/HeJ mice (five per group, per isolate) that were inoculated intradermally with 1×10^7* spirochetes. Infection was assessed at 2 and 4 weeks post-inoculation by serology and cultivation of tissues (ears, skin, joints, hearts, and/or bladders) in BSK-II containing sulfamethoxazole (0.05 mg/ml), phosphomycin (0.02 mg/ml), rifampin (0.05 mg/ml), trimethoprim (0.01 mg/ml) and amphotericin (0.0025 mg/ml). Cultures were monitored weekly by dark-field microscopy.

Generation of *Bb* infected *I. scapularis* ticks

To generate naturally-infected ticks, 300 to 400 pathogen-free *I. scapularis* larvae (Oklahoma State University) were placed on infected C3H/HeJ mice 2 to 3 weeks after needle inoculation with CE162 as described above; larvae were allowed to feed to repletion and then held in an environmental incubator until they had molted. For experiments using *Bb* isolates that were avirulent by syringe, see above, naive larvae were infected according to the immersion method described by Policastro and Schwan [28], fed on naive C3H/HeJ mice, and allowed to molt into nymphs.

Assessment of *B. burgdorferi* survival by quantitative culture and qPCR

Spirochete burdens were assessed by quantitative PCR (qPCR) using individual pools of 10 unfed and 5 replete nymphs. Total genomic DNA was isolated from surface-sterilized nymphs using the Gentra Puregene Yeast and Bacteria kit (Qiagen) according to the manufacturer's instructions. DNAs were diluted 1:10 in water and analyzed using a TaqMan-based assay for flaB ([30] and Table S1). In some experiments, the viability of spirochetes within replete nymphs (5 per strain) was assessed by semi-solid plating in pBSK medium as previously described [61].

Assessment of spirochete dissemination in ticks and transmission to mice

Groups of 10 to 12 infected unfed nymphs were confined to capsules affixed to the backs of naive C3H/HeJ mice [14]. At 72 h post-placement, approximately 5–6 feeding nymphs were forcible removed, surface-sterilized, and hemolymph collected for culturing in BSK-II as previously described [30], while the remaining ticks were allowed to feed to repletion. At 24, 48 and 72 h post-repletion, groups of mice (3–4 per strain, per time point) used as a blood meal source for infected nymphs were sacrificed and their hearts, and/or bladders) in BSK-II containing sulfamethoxazole (0.01 mg/ml) and amphotericin (0.0025 mg/ml). Survival of each specimen (i.e., from basement membrane to basement membrane, Figure S3A). As feeding progresses, influx of the blood meal and expansion of the luminal space precludes our ability to image through the full thickness (>100 μm) of individual diverticula. Consequently, optical sections of 72 h-fed midguts were acquired through a single epithelial layer (i.e., from the basement membrane to the luminal surface, Figure S3B). Representative composites of three consecutive 1-μm optical sections acquired (i) at the luminal surface; (ii) midway through the epithelium, and (iii) at the basement membrane are used to present the distribution of spirochetes through the midgut; the midway point for each specimen was determined during image processing based on the thickness of the epithelial layer. Routine optical sections of unfed and fed midguts were acquired in the coronal plane (Figure S3C). To visualize *Aptos-Gfp* organisms within 72 h-fed midguts, however, two additional steps were required to reduce light scattering and absorption by the luminal contents. First, we carefully removed a portion (~30–40%) of the blood meal to minimize interference from the blood meal and reduce the overall thickness of the midgut. Individual diverticula were then imaged in the transverse plane beginning at or near the tip (Figure S3D). A minimum of 20 *Bb*-infected nymphal midguts were examined at each time point. Epifluorescence microscopy was performed on an Olympus BX41 microscope equipped with a Retiga Exi (QImaging) camera; images were acquired using a 40× (1.4 NA) oil immersion objective with QCapture software v. 2.1 (QImaging). Fed midguts also were processed for paraffin-embedding and silver staining using the Wharthin-Starry method as previously described [30].

Transmission Electron Microscopy (TEM)

For TEM, fed midguts were isolated at 48 and 72 h post-placement and immersed for 3 h at 4°C in 0.1 M sodium cacodylate buffer (CAB) containing 4% paraformaldehyde and 2.5% glutaraldehyde. The midguts then were embedded in 1% agarose solution in 0.1 M CAC buffer. After solidification of the agarose, the midguts were sliced into >1 mm fragments, transferred to fresh 0.1 M CAB buffer and incubated overnight at 4°C. The specimens then were rinsed with fresh 0.1 M CAB buffer and post-fixed for 2 h in 0.1 M CAB buffer containing 1% OsO₄, and 0.8% potassium ferricyanide. Individual midgut slices were washed in water, stained with 1% uranyl acetate, dehydrated in ascending ethanol solutions, and embedded in PolyBed812. Thin sections (70 nm) were cut on an ultramicrotome (Leica), collected on formvar coated slot grids, and stained in 2.5% uranyl acetate and lead citrate. The stained thin sections were viewed on...
a transmission electron microscope (Hitachi H-7650) at 80 kV. Images were acquired using an AMT camera and Image Capture Engine v6.01 software (Advanced Microscopy Techniques). All images were processed using ImageJ v1.42 software [62]. In Figure 5, color overlays were added using Adobe Photoshop (CS4).

**Round body formation and recovery**

Serum starvation experiments were performed as previously described [32]. Briefly, aliquots (~400 μl) of temperature-shifted cultures at late logarithmic phase were transferred to 15 ml of RPMI-1640 (Invitrogen, catalogue #11875) at a final density of 1.5 x 10^7 spirochetes/ml and incubated at 37°C. At 24 h intervals, organisms were enumerated by darkfield microscopy and spirochete morphology was assessed by either darkfield (non-fluorescent isolates) or epifluorescence (fluorescent isolates) microscopy. To assess the ultrastructure of round bodies, spirochetes cultivated in RPMI or BSK-II for 3 days were centrifuged at 6800 g, rinsed in PBS, and resuspended in 0.1 M cac buffer containing 4% paraformaldehyde and 2.5% glutaraldehyde. The organisms were incubated on ice for 15 minutes then centrifuged at 8000 x g. Cell pellets were embedded in agarose and processed for examination by TEM as described above. In vitro generated round bodies were recovered by the addition of an equal volume of BSK-II after 4 days of cultivation in RPMI. Recovery was monitored in real-time using StreamPix software (Norpix) on an Olympus BX41 microscope equipped with a Retiga Exi camera. Videos were generated using QuickTime Player v.7.7 (Apple, Inc.). The recovery of round bodies in nymphs at 72 h post-placement was performed by first dissecting the midgut in RPMI. The diverticula were then detached from the stomach using a scalpel blade. A cover slip was then applied to the cover glass and real-time imaging was performed for the RPMI control. Round body recovery in nymphal midguts was induced by adding 10 μl BSK-II to the dissected midguts. Recovery was monitored in real-time as described above.

**Bioinformatics**

Candidate surface-exposed lipoproteins within the RpoS regulon [13] were identified using the database created by Setubal et al. [53]. Non-surface exposed lipoproteins then were excluded based on published data [49,63,64]. Candidate RpoS-dependent β-barrel-outer membrane spanning proteins (OMPs) were identified and filtered using the consensus computational framework recently used to identify OMPs in *Treponema pallidum* [54]. The proteins identified by two or more OMP localization/topology programs were designated OMP candidates (Table S3).

**Statistical analysis**

Statistical analyses were performed on qPCR, spirochete viability, and round body formation using Prism 5 (GraphPad Software, Inc.). Student’s t tests were used to calculate the significance of the qPCR and spirochete viability (p≤0.05 was considered significant) and is indicated in the figures and/or legends. Unpaired student’s t test with two-tailed p values were used to compare round body formation between different Bb isolates at each of the time points studied. For qRT-PCR, the normalized copy number values for each gene of interest were compared in Prism using an unpaired t-test with two-tailed p values and a 95% confidence interval.

**Supporting Information**

**Figure S1** Spirochete persistence and survival in *I. scapularis* are not affected by loss of either RpoS or OspC. (A) Spirochete burdens of WT, ΔrpoS, RpoS complemented ΔrpoS (Compl) and ΔospC before and after feeding on naive C3H/HeJ mice. The nymphs used in these experiments were used in Table 2. WT-gfp and ΔrpoS-gfp (B) burdens determined by qPCR and (C) viability, assessed by semi-solid plating, are highly similar in unfed (black) and fed (gray) nymphs. Values represent the means ± SEMs from three independent experiments. (TIF)

**Figure S2** Contours of the RpoSBb regulon in *I. scapularis*. qRT-PCR analysis of (A) absolutely and (B) partially RpoS-dependent upregulated genes selected from microarray data derived from Bb cultivated within DMCs [13]. Expression profiling was performed using fed larvae, unfed and fed nymphs that had been naturally-infected with WT Bb as well as fed nymphs that had been infected as larvae by immersion with either WT-gfp or ΔrpoS-gfp isolates. Values represent the average β-lactamase-normalized transcript copy number ± SEM for each gene; average values are considered significantly different when p is ≤0.05 (indicated by asterisks). (TIF)

**Figure S3** Cartoon illustrating the scheme used to acquire optical sections through unfed and fed nymphal midguts isolated at 48 and 72 h post-placement. Serial Z-series confocal images were generated by obtaining 1-μm optical sections through the depth of unfed and fed midguts as described in Materials and Methods. (A) Cartoon depicting the scheme used to image individual diverticulum from unfed and 48 h-fed nymphal midguts; for these types of specimens, optical sections were acquired in the coronal plane through the full thickness of a midgut. (B) Cartoon depicting the scheme used to acquire images of an individual diverticulum from a 72 h-fed midgut in the coronal and transverse planes. (C) Zoom-in of coronal views for 72 h-fed nymphal midguts infected with either WT or ΔrpoS organisms illustrating the depths at which fluorescent spirochetes can be visualized by confocal microscopy. (D) Zoom-in of 72 h-fed nymphal midguts infected with ΔrpoS organisms imaged in the transverse plane after a portion of the blood meal had been removed; note the difference in the luminal space in the coronal and transverse planes. Solid-line arrows are used to indicate the laser beam, while a dashed line and “eye” are used to represent what is detected by the microscope’s photomultiplier tube. Abbreviations: LS, luminal surface; MW, midway through the epithelial layer; and BM, basement membrane. (TIF)

**Figure S4** Spirochetes introduced by immersion undergo biphasic dissemination. (A) Spirochete burdens are identical in unfed and fed nymphs that acquired WT Bb as larvae by immersion or naturally by feeding on infected mice. Burdens were determined by qPCR; values represent the means ± SDs from three independent experiments. (B) The distribution of WT-gfp spirochetes in nymphs infected as larvae by immersion is highly similar to that of spirochetes within naturally-infected nymphs [30]. Composite images depicting midguts of nymphs infected by immersion are the same as those presented in Figures 2 and 3, while the images of midguts isolated from naturally-infected nymphs were taken from [30] and used with permission. The leftmost images in each panel “Full Composite” depict the full thickness of the midgut, while 3-μm composite images show spirochetes at the luminal surface, midway through the epithelial layer, and at the basement membrane. A detailed schematic indicating how confocal images of unfed and fed midguts were acquired is presented in Figure S9. Green
represents GFP+ spirochetes and red indicates midgut epithelial cells labeled with FM4-64; scale bars = 25 μm. Images of naturally infected nymphs are reproduced with permission from Live imaging reveals a biphasic mode of dissemination of Borrelia burgdorferi within ticks, published in Volume 119, Issue 12 (December 1, 2009), J Clin Invest, 2009; 119(12):3652-3665. doi:10.1172/JCI39401. Copyright © 2009, American Society for Clinical Investigation [30].

Figure S5  Complementation of ΔrpoS Bb with a wild-type copy of rpoS restores expression of RpoS-dependent genes following temperature-shift. Whole cell B. burgdorferi lysates were prepared from WT (Bb914), ΔrpoS (Bb1058) and RpoS-complemented (SE186) isolates, separated by SDS-PAGE and stained with silver as described in Materials and Methods. (TIF)

Figure S6  ArpoS organisms have an altered morphology and distribution pattern within 72 h-fed midguts. Representative composite images of cryosectioned midguts infected with (A) WT-gfp or (B) ArpoS-gfp Bb; green represents spirochetes expressing GFP while red indicates midgut epithelial cells labeled with FM4-64; scale bars = 25 μm. (TIF)

Figure S7  Loss of RpoS and CoaDR enhances round body formation under nutrient-limiting conditions in vitro. Representative images of (A) WT-gfp, (B) ArpoS-gfp, (C) complemented ΔrpoS-gfp, (D) ΔcoaDR, and (E) complemented ΔcoaDR isolates after 3 days in RPMI; scale bar = 50 μm. (TIF)

Table S1  Oligonucleotides used in this study. (DOC)
Table S2  Virulence of B. burgdorferi strains following needle-inoculation. (DOC)
Table S3  Known and/or putative RpoS-dependent tick midgut adhesins. (DOC)

Video S1  Round bodies generated in vitro rapidly elongate into spirochetes. Spirochetes were transformed into round bodies by culturing either WT-gfp or ArpoS-gfp isolates in RPMI for 4 days. The video is arranged into two segments depicting the rapid transformation of WT-gfp or ΔrpoS-gfp round bodies into normal spirochetes after the addition of Bsk-II (see the upper left hand corner for the time stamp). (MOV)

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Author Contributions
Conceived and designed the experiments: SMDE MJC CHE JDR. Performed the experiments: SMDE MJC CHE. Analyzed the data: SMDE MJC CHE JDR. Contributed reagents/materials/analysis tools: SMDE MJC CHE JDR. Wrote the paper: SMDE MJC JDR.

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